

## Anti-inflammatory cannabinoids in diet

Towards a better understanding of CB<sub>2</sub> receptor action?

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### Abstract

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The endocannabinoid system is an ancient lipid signaling network which in mammals modulates neuronal functions, inflammatory processes, and is involved in the aetiology of certain human lifestyle diseases, such as Crohn's disease, atherosclerosis and osteoarthritis. The system is able to downregulate stress-related signals that lead to chronic inflammation and certain types of pain, but it is also involved in causing inflammation-associated symptoms, depending on the physiological context. The cannabinoid type-2 (CB<sub>2</sub>) receptor, which unlike the CB<sub>1</sub> receptor does not induce central side effects, has been shown to be a promising therapeutic target. While CB<sub>1</sub> receptor antagonists/inverse agonists are of therapeutic value, also CB<sub>2</sub> receptor ligands including agonists are of pharmacological interest. Although the endocannabinoid system is known to be involved in the regulation of energy homeostasis and metabolism (mainly via CB<sub>1</sub> receptors) there was hitherto no direct link between food intake and cannabinoid receptor activation. Our recent finding that beta-caryophyllene, a ubiquitous lipophilic plant natural product, selectively binds to the CB<sub>2</sub> receptor and acts as a full agonist is unexpected. Maybe even more unexpected is that oral administration of this dietary compound exerts potent anti-inflammatory effects in wild type mice but not in CB<sub>2</sub> receptor (Cnr2<sup>-/-</sup>) knockout mice. Like other CB<sub>2</sub> ligands also beta-caryophyllene inhibits the pathways triggered by activation of the toll-like receptor complex CD14/TLR4/MD2, which typically lead to the expression of proinflammatory cytokines (IL-1 $\beta$ , IL-6; IL-8 and TNF $\alpha$ ) and promotes a TH<sub>1</sub> immune response. In this addendum, the CB<sub>2</sub> receptor-dependent effect of beta-caryophyllene on LPS-triggered activation of the kinases Erk1/2 and JNK1/2 are further discussed with respect to the possibility that both CB<sub>2</sub> inverse agonists and agonists, independent of their G-protein signaling, may block LPS-triggered activation of MAPKs, leading to inhibition of proinflammatory cytokine expression and attenuation of inflammation.

**Keywords:** cannabinoid CB<sub>2</sub> receptor, MAPK, TLR4, TNF $\alpha$ , inflammation, caryophyllene

Apart from the well-known psychomimetic action of the classical cannabinoid  $\Delta^9$ -tetrahydrocannabinol (THC) mediated by centrally expressed cannabinoid CB<sub>1</sub> receptors, it is now generally accepted that the endocannabinoid system also occupies regulatory roles in peripheral tissues.<sup>1-3</sup> The cannabinoid CB<sub>2</sub> receptor, which is involved in the tuning of

inflammatory processes,<sup>4-6</sup> is able to mediate cellular signals that in many cases lead to attenuation of inflammation.<sup>7,8</sup> The endogenous CB receptor ligands, the arachidonic acid derivatives anandamide and 2-arachidonoylglycerol (2-AG), which are the major endocannabinoids known, partially or fully activate CB receptors in a rather non-selective manner and controlled by strict biosynthesis and degradation.<sup>9</sup>

Anandamide and 2-AG have been shown to inhibit the inflammatory processes triggered upon activation of the toll-like receptor complex CD14/TLR4/MD2 (i.e., LPS and carrageenan stimulation).<sup>10,11</sup> The same effect has also been reported for other CB<sub>2</sub> receptor agonists like JWH133,<sup>12</sup> HU-308,<sup>13</sup> and *N*-alkylamides.<sup>14</sup> Paradoxically, also CB<sub>2</sub> receptor inverse agonists like JTE-907,<sup>15</sup> SR144528,<sup>15,16</sup> and Sch.336,<sup>17</sup> show this effect in the same or similar models. As both CB<sub>2</sub> receptor inverse agonists<sup>18</sup> and agonists<sup>7</sup> block identical pathways in an apparently CB<sub>2</sub> receptor-dependent manner, mediators other than cAMP or calcium transients should be involved in this mechanism. In our study,<sup>19</sup> treatment of primary monocytes/macrophages with the CB<sub>2</sub> receptor agonist beta-caryophyllene leads to inhibition of LPS-stimulated phosphorylation of the kinases Erk1/2 and JNK1/2 but not p38, despite the fact that the phosphorylation of these kinases is increased by this compound and JWH133 in resting (non-stimulated) monocytes/macrophages. Thus, the context of differentially activated signaling pathways determines the way signals are integrated. Erk1/2 has been shown to be either activated or inhibited by different GPCRs, both via G-protein dependent and independent mechanisms, involving intracellular calcium transients,<sup>20</sup> the  $\beta$ -arrestin scaffold,<sup>21</sup> and ubiquitination.<sup>22</sup> JNK1/2 is a stress-activated protein kinase that can be modulated by GPCRs. Protein tyrosine kinases are thought to be involved in the regulation of signal transmission from GPCRs to activation of the JNK/SAPK kinase pathway.<sup>23</sup> To inhibit LPS-stimulated Erk1/2 and JNK phosphorylation (i.e., inhibition of their pathways) could be achieved by blocking MyD88, TRAF6 and IKK $\gamma$ , which would lead to reduced transcription of proinflammatory cytokines and metalloproteinases. The question remains how CB<sub>2</sub> receptor ligands as diverse as inverse agonists and full agonists do the same thing in an apparently receptor-dependent fashion and whether in all cases MAPKs are involved. A direct inhibition of NF $\kappa$ B, which is the primary signal in LPS-induced gene expression, is more unlikely because this factor is inhibited only at higher  $\mu$ M-concentrations of cannabinoids. Thus, how exactly does the CB<sub>2</sub> receptor modulate MAPKs in immune cells? There are different possible mechanisms, such as hitherto neglected CB<sub>2</sub> pathways like the ras-MAPK pathway or even an indirect action via TNF $\alpha$  expression. Interestingly, accumulation of TNF $\alpha$  transcripts, but inhibition of protein expression has been shown after treatment with both CB<sub>2</sub> selective inverse agonists and agonists.<sup>24,25</sup> On the same line, inhibition of the JNK pathway in monocytes/macrophages will inhibit TNF $\alpha$  translation.<sup>26</sup> Moreover, TNF $\alpha$  stimulated pathways could be inhibited indirectly, such that the pro-inflammatory feedback will shut down.

In fact, there is good evidence that constitutive TNF $\alpha$  is inhibited by the endocannabinoid system and that the CB<sub>2</sub> receptor is involved.<sup>27,28</sup> Moreover, TNF $\alpha$  can activate the endocannabinoid system in adipose tissue.<sup>29</sup> As TNF $\alpha$  triggers the activation of Ras, p38 MAPK, ERK1/2, SAPK/JNK and Akt pathways and ultimately proinflammatory cytokine expression and cellular proliferation and migration, it is also possible that the autocrine action of TNF $\alpha$  may be inhibited by cannabinoids. The CB<sub>2</sub> agonists JWH-133 and HU-308 dose-dependently attenuate the effects of TNF $\alpha$ .<sup>30,31</sup> On the other hand, activation of the CB<sub>2</sub> receptor by JWH133 in macrophages has been shown to activate Erk1/2 which leads to expression of IL-10 and thus a counteraction to proinflammatory gene expression.<sup>32</sup> The CB<sub>2</sub>receptor has also been found to be a potential target to prevent atherosclerosis,<sup>33</sup> in part via TNF $\alpha$  signalling.<sup>34</sup> Because CB<sub>2</sub> receptor activation attenuates TNF $\alpha$ -induced endothelial cell activation, transendothelial migration of monocytes, and monocyte/neutrophil-endothelial adhesion, and decreases TNF $\alpha$ -induced proliferation and migration of human coronary vascular smooth muscle cells,<sup>35</sup> in addition to inhibiting constitutive and LPS-stimulated TNF $\alpha$  expression, the CB<sub>2</sub> receptor may be an essential pleiotropic modulator of TNF $\alpha$  signal transduction. Providing that the multifunctional cytokine TNF $\alpha$  is involved in numerous pathophysiological processes and the endocannabinoid system can dynamically regulate the levels and effects of this cytokine, more research should be dedicated to the molecular understanding of CB receptor signaling in the immune system.

Unfortunately, the pharmacological tool box for CB<sub>2</sub> receptors is still very limited, the results with selective agonists and antagonists often confusing and the conclusions drawn from such experiments potentially erroneous. For example, it would be wrong to say that 2-AG does mediate intracellular calcium in HL60 cells in a CB<sub>2</sub> receptor-independent manner because the selective CB<sub>2</sub>antagonist/inverse agonist AM630 cannot block this effect. The CB<sub>2</sub> inverse agonist SR144528 can exactly do this.<sup>36</sup> In some experiments AM630 and SR144528 are able to block the anti-inflammatory effects of CB<sub>2</sub> agonists; in other experiments SR144528 is anti-inflammatory. Thus, the classical concept of agonists and antagonists, “one lock and one key”, does not sufficiently describe cannabinoid GPCR signaling. Even more puzzling is that CB<sub>2</sub> receptor-selective agonists like HU308 increase the inflammation caused by allergens in the skin, while CB<sub>2</sub> antagonists and CB<sub>1</sub> agonists inhibit inflammation,<sup>37</sup> thus pointing to the possibility that different cell types mediate distinct signals. The apparent involvement of CBreceptors and Erk1/2 and JNK MAPKs in the production of inflammatory signals and skin cancer development after UVB irradiation seems to confirm this.<sup>38</sup>

Even though we do obviously not yet understand the underlying molecular mechanisms of these observations, it can be said that in most cases systemic administration of CB<sub>2</sub> receptor-selective ligands leads to attenuation of systemic inflammation in different animal models. It needs to be emphasized, however, that it is currently impossible to determine the molecular mechanism of a ligand in vivo and whether agonists are doing what

antagonists should do and vice versa. One hint in that direction comes from data showing that the CB<sub>1</sub> antagonist SR141716A acts like a partial agonist in vivo.<sup>39</sup>

In the case of the dietary natural product beta-caryophyllene, a full CB<sub>2</sub> receptor-selective agonist in vitro, potent anti-inflammatory cannabimimetic effects are observed.<sup>19</sup> Intriguingly, the lowest oral dose tested (5 mg/Kg) of this widespread and apparently non-toxic compound, which is also an FDA-approved food additive, was the most effective. Maybe this strengthens the hypothesis that beta-caryophyllene is indeed a dietary cannabinoid, thus inferring that by eating this compound the endocannabinoid system may be modulated in a beneficial way, possibly through modulation of TNF $\alpha$  pathways, maybe even to such a degree that certain lifestyle diseases could be prevented. The finding that oral beta-caryophyllene is more effective in mice than JWH-133 despite its significantly lower CB<sub>2</sub> receptor affinity is encouraging. However, further studies will have to determine whether this natural product is also bioavailable in humans and in more general terms, how CB<sub>2</sub> receptor ligands exactly modulate inflammation in vivo.

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## Notes

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## Footnotes

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## References

1. Di Marzo V. Targeting the endocannabinoid system: to enhance or reduce? Nat Rev Drug Discov. 2008;7:438–455. [[PubMed](#)]
2. Kunos G, Osei-Hyamian D. Endocannabinoids and liver disease IV. Endocannabinoid involvement in obesity and hepatic steatosis. Am J Physiol Gastrointest Liver Physiol. 2008;294:1101–1104. [[PubMed](#)]
3. Klein TW, Newton CA. Therapeutic potential of cannabinoid-based drugs. Adv Exp Med Biol. 2007;601:395–413. [[PubMed](#)]
4. Wolf SA, Ullrich O. Endocannabinoids and the brain immune system: new neurones at the horizon? J Neuroendocrinol. 2008;20:15–19. [[PubMed](#)]
5. Wright KL, Duncan M, Sharkey KA. Cannabinoid CB<sub>2</sub> receptors in the gastrointestinal tract: a regulatory system in states of inflammation. Br J Pharmacol. 2008;153:263–270. [[PMC free article](#)][[PubMed](#)]

6. Lotersztajn S, Teixeira-Clerc F, Julien B, Deveaux V, Ichigotani Y, Manin S, Tran-Van-Nhieu J, Karsak M, Zimmer A, Mallat A. CB<sub>2</sub> receptors as new therapeutic targets for liver diseases. *Br J Pharmacol*. 2008;153:286–289. [[PMC free article](#)] [[PubMed](#)]
7. Ashton JC. Cannabinoids for the treatment of inflammation. *Curr Opin Investig Drugs*. 2007;8:373–384. [[PubMed](#)]
8. Guindon J, Hohmann AG. Cannabinoid CB<sub>2</sub> receptors: a therapeutic target for the treatment of inflammatory and neuropathic pain. *Br J Pharmacol*. 2008;153:319–334. [[PMC free article](#)] [[PubMed](#)]
9. Di Marzo V. Endocannabinoids: synthesis and degradation. *Rev Physiol Biochem Pharmacol*. 2008;160:1–24. [[PubMed](#)]
10. Klegeris A, Bissonnette CJ, McGeer PL. Reduction of human monocytic cell neurotoxicity and cytokine secretion by ligands of the cannabinoid-type CB<sub>2</sub> receptor. *Br J Pharmacol*. 2003;139:775–786. [[PMC free article](#)] [[PubMed](#)]
11. Gallily R, Breuer A, Mechoulam R. 2-Arachidonylglycerol, an endogenous cannabinoid, inhibits tumor necrosis factor- $\alpha$  production in murine macrophages, and in mice. *Eur J Pharmacol*. 2000;406:5–7. [[PubMed](#)]
12. Duncan M, Mouihate A, Mackie K, Keenan CM, Buckley NE, Davison JS, Patel KD, Pittman QJ, Sharkey KA. Cannabinoid CB<sub>2</sub> receptors in the enteric nervous system modulate gastrointestinal contractility in lipopolysaccharide-treated rats. *Am J Physiol Gastrointest Liver Physiol*. 2008 Ahead of print.
13. Hanus L, Breuer A, Tchilibon S, Shiloah S, Goldenberg D, Horowitz M, Pertwee RG, Ross RA, Mechoulam R, Fride E. HU-308: a specific agonist for CB(2), a peripheral cannabinoid receptor. *Proc Natl Acad Sci USA*. 1999;96:14228–14233. [[PMC free article](#)] [[PubMed](#)]
14. Raduner S, Majewska A, Chen JZ, Xie XQ, Hamon J, Faller B, Altmann KH, Gertsch J. Alkylamides from Echinacea are a new class of cannabinomimetics. Cannabinoid type 2 receptor-dependent and -independent immunomodulatory effects. *J Biol Chem*. 2006;281:14192–14206. [[PubMed](#)]
15. Ueda Y, Miyagawa N, Matsui T, Kaya T, Iwamura H. Involvement of cannabinoid CB(2) receptor-mediated response and efficacy of cannabinoid CB(2) receptor inverse agonist, JTE-907, in cutaneous inflammation in mice. *Eur J Pharmacol*. 2005;520:164–171. [[PubMed](#)]
16. Oka S, Yanagimoto S, Ikeda S, Gokoh M, Kishimoto S, Waku K, Ishima Y, Sugiura T. Evidence for the involvement of the cannabinoid CB<sub>2</sub> receptor and its endogenous ligand 2-arachidonoylglycerol in 12-O-tetradecanoylphorbol-13-acetate-induced acute inflammation in mouse ear. *J Biol Chem*. 2005;280:18488–18497. [[PubMed](#)]

17. Lunn CA, Fine JS, Rojas-Triana A, Jackson JV, Fan X, Kung TT, Gonsiorek W, Schwarz MA, Lavey B, Kozlowski JA, Narula SK, Lundell DJ, Hipkin RW, Bober LA. A novel cannabinoid peripheral cannabinoid receptor-selective inverse agonist blocks leukocyte recruitment in vivo. *J Pharmacol Exp Ther.* 2006;316:780–788. [[PubMed](#)]
18. Lunn CA, Reich EP, Fine JS, Lavey B, Kozlowski JA, Hipkin RW, Lundell DJ, Bober L. Biology and therapeutic potential of cannabinoid CB<sub>2</sub> receptor inverse agonists. *Br J Pharmacol.* 2008;153:226–239. [[PMC free article](#)] [[PubMed](#)]
19. Gertsch J, Leonti M, Raduner S, Racz I, Chen JZ, Xie XQ, Altmann KH, Karsak M, Zimmer A. Beta-caryophyllene is a dietary cannabinoid. *Proc Natl Acad Sci USA.* 2008 Ahead of Print.
20. Chuderland D, Marmor G, Shainskaya A, Seger R. Calcium-mediated interactions regulate the subcellular localization of extracellular signal-regulated kinases. *J Biol Chem.* 2008;283:11176–11188. [[PubMed](#)]
21. Gripenrog JM, Miettinen HM. Formyl peptide receptor-mediated ERK1/2 activation occurs through G(i) and is not dependent on beta-arrestin1/2. *Cell Signal.* 2008;20:424–431. [[PMC free article](#)][[PubMed](#)]
22. Shenoy SK, Lefkowitz RJ. Receptor-specific ubiquitination of beta-arrestin directs assembly and targeting of seven-transmembrane receptor signalosomes. *J Biol Chem.* 2005;280:15315–15324. [[PubMed](#)]
23. Dikic I, Blaukat A. Protein tyrosine kinase-mediated pathways in G protein-coupled receptor signaling. *Cell Biochem Biophys.* 1999;30:369–387. [[PubMed](#)]
24. Puffenbarger RA, Boothe AC, Cabral GA. Cannabinoids inhibit LPS-inducible cytokine mRNA expression in rat microglial cells. *Glia.* 2000;29:58–69. [[PubMed](#)]
25. Gertsch J, Schoop R, Kuenzle U, Suter A. Echinacea alkylamides modulate TNFalpha gene expression via cannabinoid receptor CB<sub>2</sub> and multiple signal transduction pathways. *FEBS Lett.* 2004;577:563–569. [[PubMed](#)]
26. Swantek JL, Cobb MH, Geppert TD. Jun N-terminal kinase/stress-activated protein kinase (JNK/SAPK) is required for lipopolysaccharide stimulation of tumor necrosis factor alpha (TNFalpha) translation: glucocorticoids inhibit TNFalpha translation by blocking JNK/SAPK. *Mol Cell Biol.* 1997;17:6274–6282. [[PMC free article](#)] [[PubMed](#)]
27. Rajesh M, Mukhopadhyay P, Haskó G, Huffman JW, Mackie K, Pacher P. CB<sub>2</sub> cannabinoid receptor agonists attenuate TNFalpha-induced human vascular smooth muscle cell proliferation and migration. *Br J Pharmacol.* 2008;153:347–357. [[PMC free article](#)] [[PubMed](#)]

28. Mechoulam R, Shohami E. Endocannabinoids and traumatic brain injury. *Mol Neurobiol.* 2007;36:68–74. [[PubMed](#)]
29. Kempf K, Hector J, Strate T, Schwarzloh B, Rose B, Herder C, Martin S, Algenstaedt P. Immune-mediated activation of the endocannabinoid system in visceral adipose tissue in obesity. *Horm Metab Res.* 2007;39:596–600. [[PubMed](#)]
30. Bátkai S, Osei-Hyiaman D, Pan H, El-Assal O, Rajesh M, Mukhopadhyay P, Hong F, Harvey-White J, Jafri A, Haskó G, Huffman JW, Gao B, Kunos G, Pacher P. Cannabinoid-2 receptor mediates protection against hepatic ischemia/reperfusion injury. *FASEB J.* 2007;21:1788–1800. [[PMC free article](#)] [[PubMed](#)]
31. Rajesh M, Pan H, Mukhopadhyay P, Bátkai S, Osei-Hyiaman D, Haskó G, Liaudet L, Gao B, Pacher P. Cannabinoid-2 receptor agonist HU-308 protects against hepatic ischemia/reperfusion injury by attenuating oxidative stress, inflammatory response, and apoptosis. *J Leukoc Biol.* 2007;82:1382–1389. [[PMC free article](#)] [[PubMed](#)]
32. Correa F, Mestre L, Docagne F, Guaza C. Activation of cannabinoid CB<sub>2</sub> receptor negatively regulates IL-12p40 production in murine macrophages: role of IL-10 and ERK1/2 kinase signaling. *Br J Pharmacol.* 2005;145:441–448. [[PMC free article](#)] [[PubMed](#)]
33. Steffens S, Veillard NR, Arnaud C, Pelli G, Burger F, Staub C, Karsak M, Zimmer A, Frossard JL, Mach F. Low dose oral cannabinoid therapy reduces progression of atherosclerosis in mice. *Nature.* 2005;434:782–786. [[PubMed](#)]
34. Pacher P, Ungvári Z. Pleiotropic effects of the CB<sub>2</sub> cannabinoid receptor activation on human monocyte migration: implications for atherosclerosis and inflammatory diseases. *Am J Physiol Heart Circ Physiol.* 2008;294:1133–1134. [[PMC free article](#)] [[PubMed](#)]
35. Rajesh M, Mukhopadhyay P, Bátkai S, Haskó G, Liaudet L, Huffman JW, Csiszar A, Ungvari Z, Mackie K, Chatterjee S, Pacher P. CB<sub>2</sub>-receptor stimulation attenuates TNF $\alpha$ -induced human endothelial cell activation, transendothelial migration of monocytes, and monocyte-endothelial adhesion. *Am J Physiol Heart Circ Physiol.* 2007;293:2210–2218. [[PMC free article](#)] [[PubMed](#)]
36. Sugiura T, Kondo S, Kishimoto S, Miyashita T, Nakane S, Kodaka T, Suhara Y, Takayama H, Waku K. Evidence that 2-arachidonoylglycerol but not N-palmitoylethanolamine or anandamide is the physiological ligand for the cannabinoid CB<sub>2</sub> receptor. Comparison of the agonistic activities of various cannabinoid receptor ligands in HL-60 cells. *J Biol Chem.* 2000;275:605–612. [[PubMed](#)]
37. Karsak M, Gaffal E, Date R, Wang-Eckhardt L, Rehnelt J, Petrosino S, Starowicz K, Steuder R, Schlicker E, Cravatt B, Mechoulam R, Buettner R, Werner S, Di Marzo V, Tüting

T, Zimmer A. Attenuation of allergic contact dermatitis through the endocannabinoid system. *Science*.2007;316:1494–1497. [[PubMed](#)]

38. Zheng D, Bode AM, Zhao Q, Cho YY, Zhu F, Ma WY, Dong Z. The cannabinoid receptors are required for ultraviolet-induced inflammation and skin cancer development. *Cancer Res*.2008;68:3992–3998. [[PMC free article](#)] [[PubMed](#)]

39. Smith SR, Terminelli C, Denhardt G. Effects of cannabinoid receptor agonist and antagonist ligands on production of inflammatory cytokines and anti-inflammatory interleukin-10 in endotoxemic mice. *J Pharmacol Exp Ther*. 2000;293:136–150. [[PubMed](#)]